

# COVID-19 : une maladie immunologique ? Réponse cellulaire au SARS-CoV-2



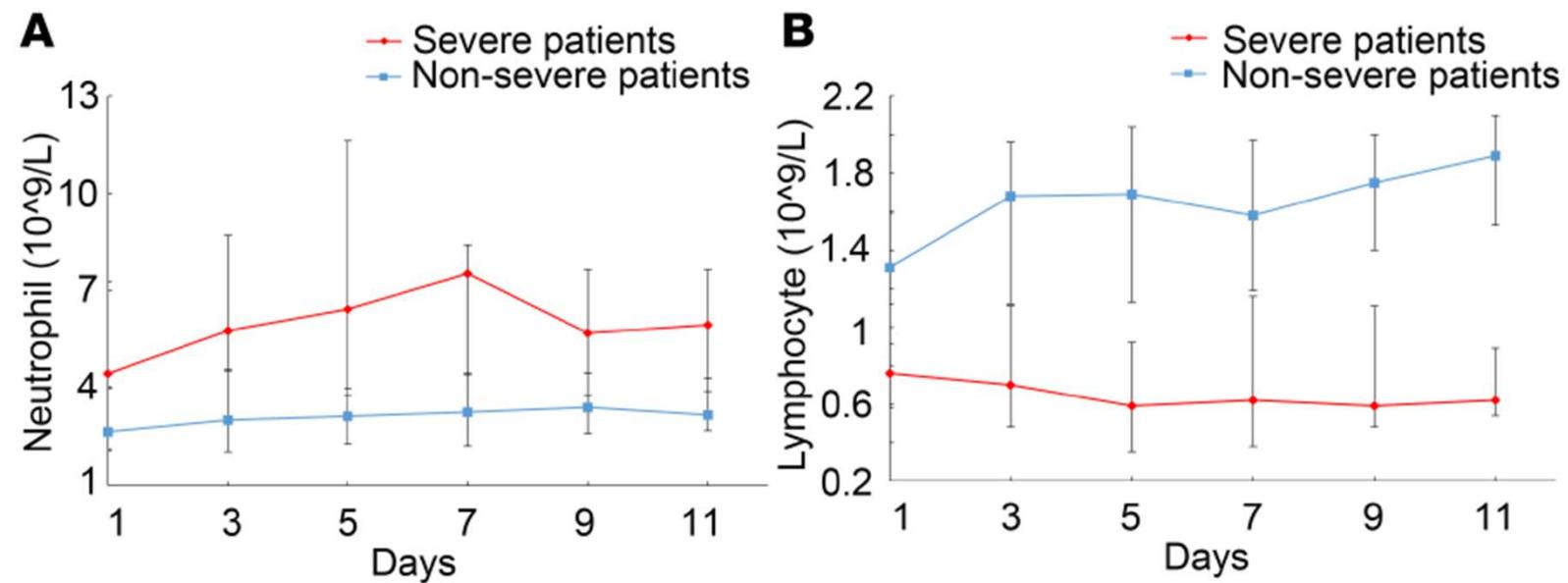
4<sup>es</sup> Journées de la Fédération d'Immunologie Médicale

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# Hyperneutrophilia and Lymphopenia during severe COVID-19 infection

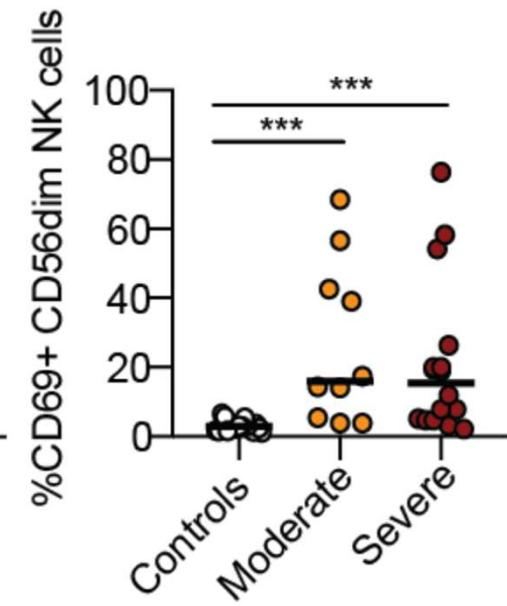
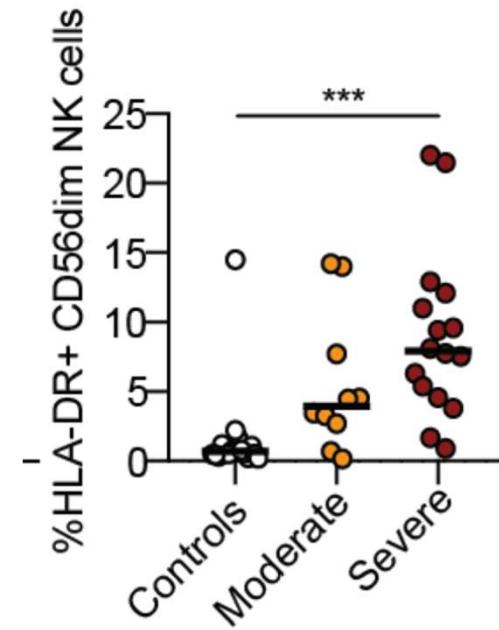
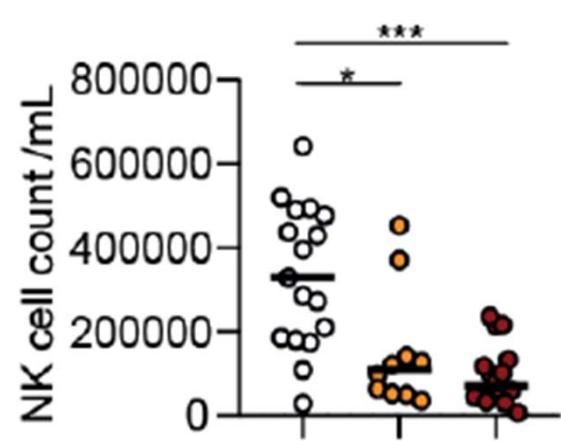
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Shaohua et al, JCI Insight 2020

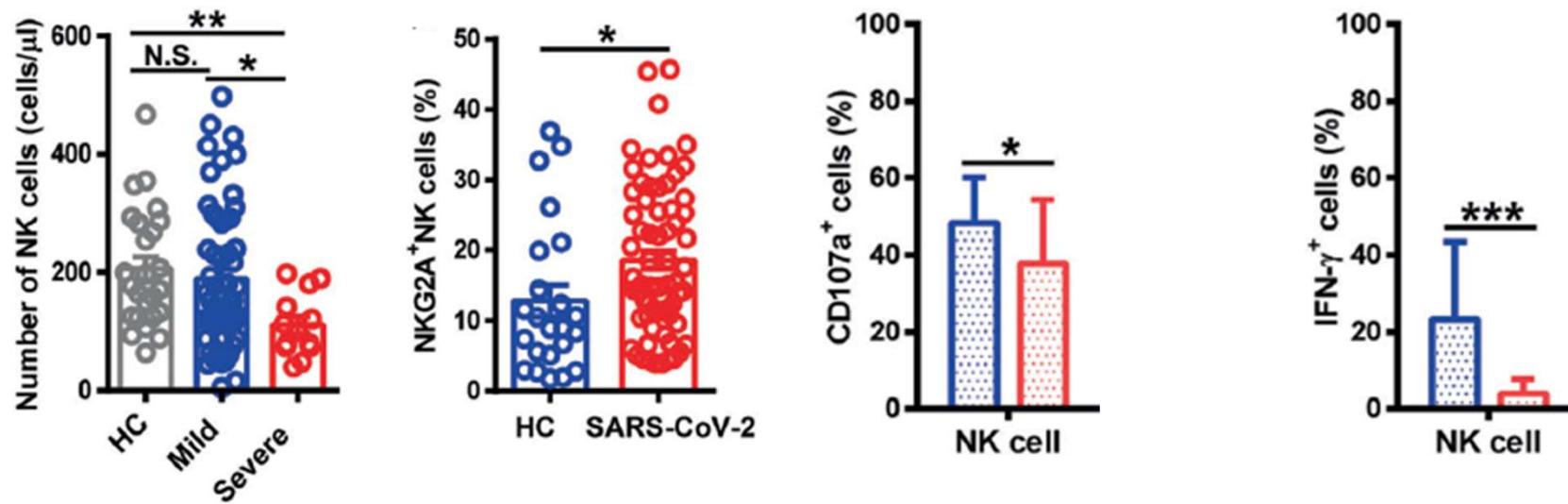
# NK cells are activated during moderate and severe COVID-19 infection

Severe (n=17)  
Moderate (n=10)  
Healthy (n=17)



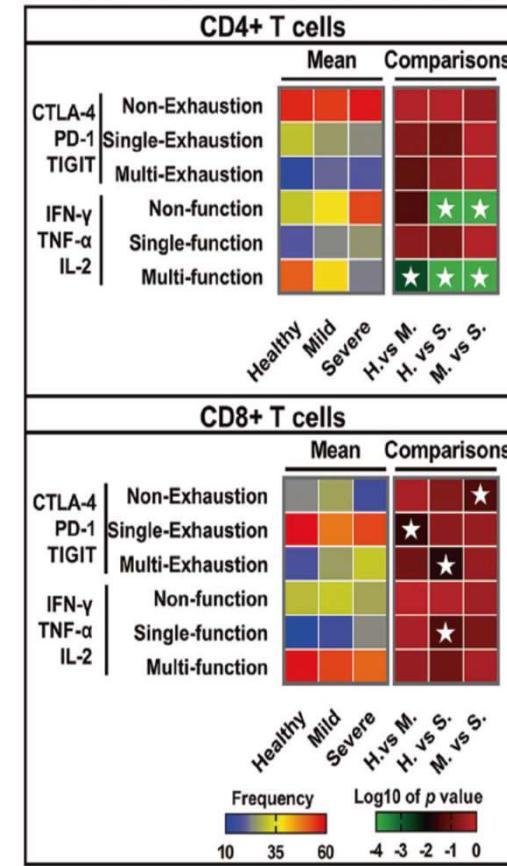
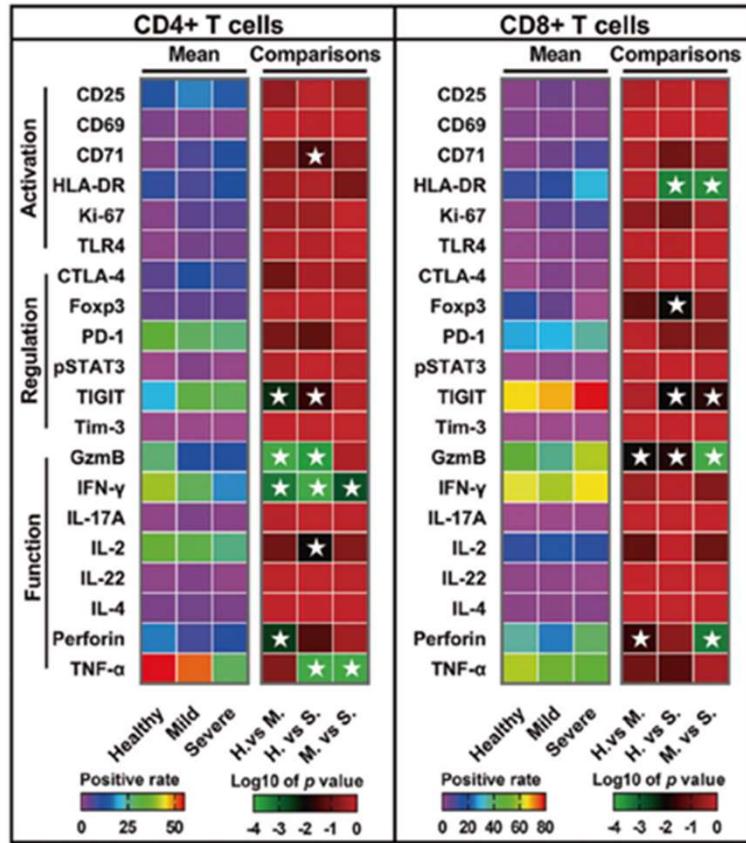
# NK cells shows an hypofunctionnal phenotype during severe COVID-19 infection

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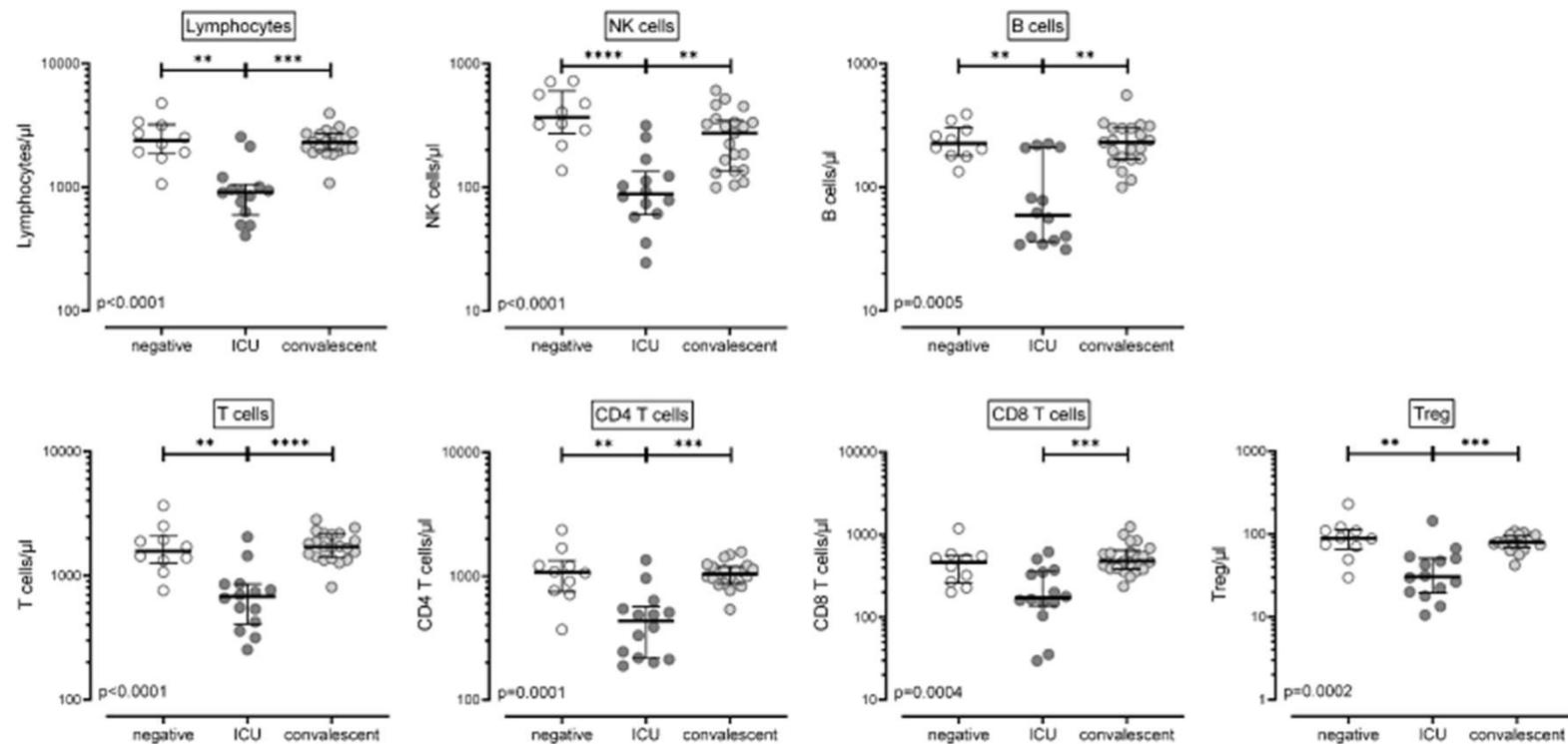
Zheng et al, Cell Mol Immunol 2020

# Peripheral CD4 T cell functions are lacking during mild/severe COVID-19 infection



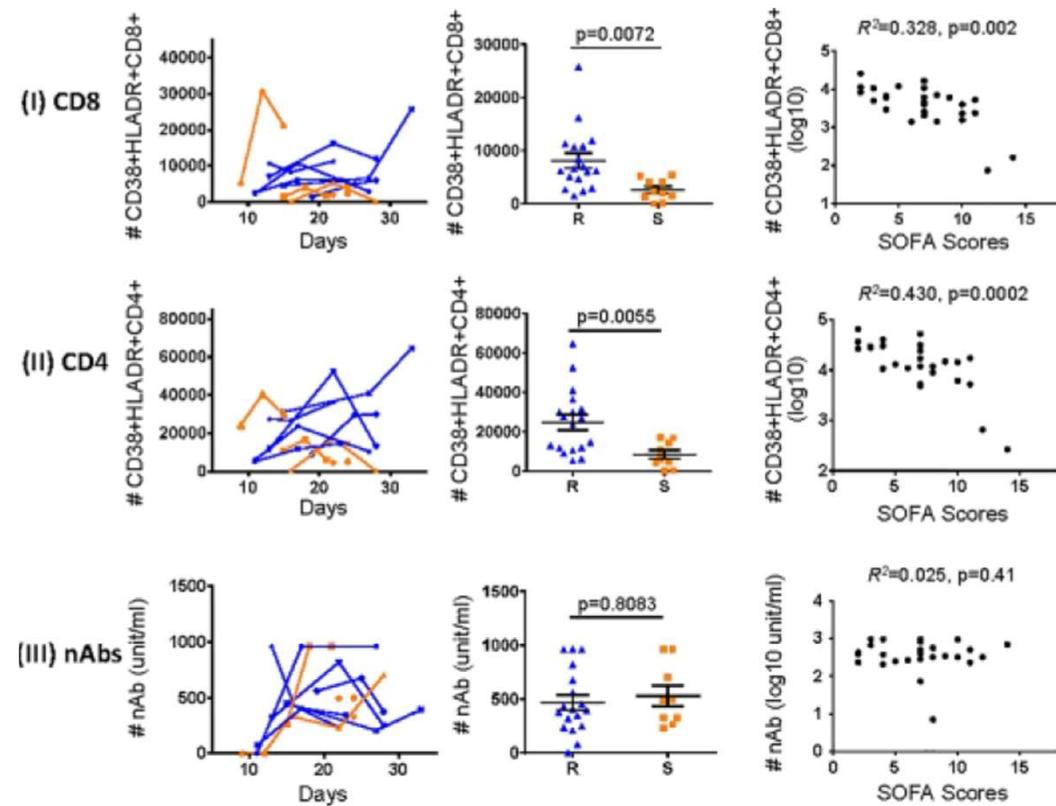
Zheng et al, Cell and Mol Immunol, 2020

# The lymphopenia is reversible after severe COVID-19 infection



Schub et al, JCI Insight 2020

# T cell activation is associated with recovery after mild COVID-19 infection

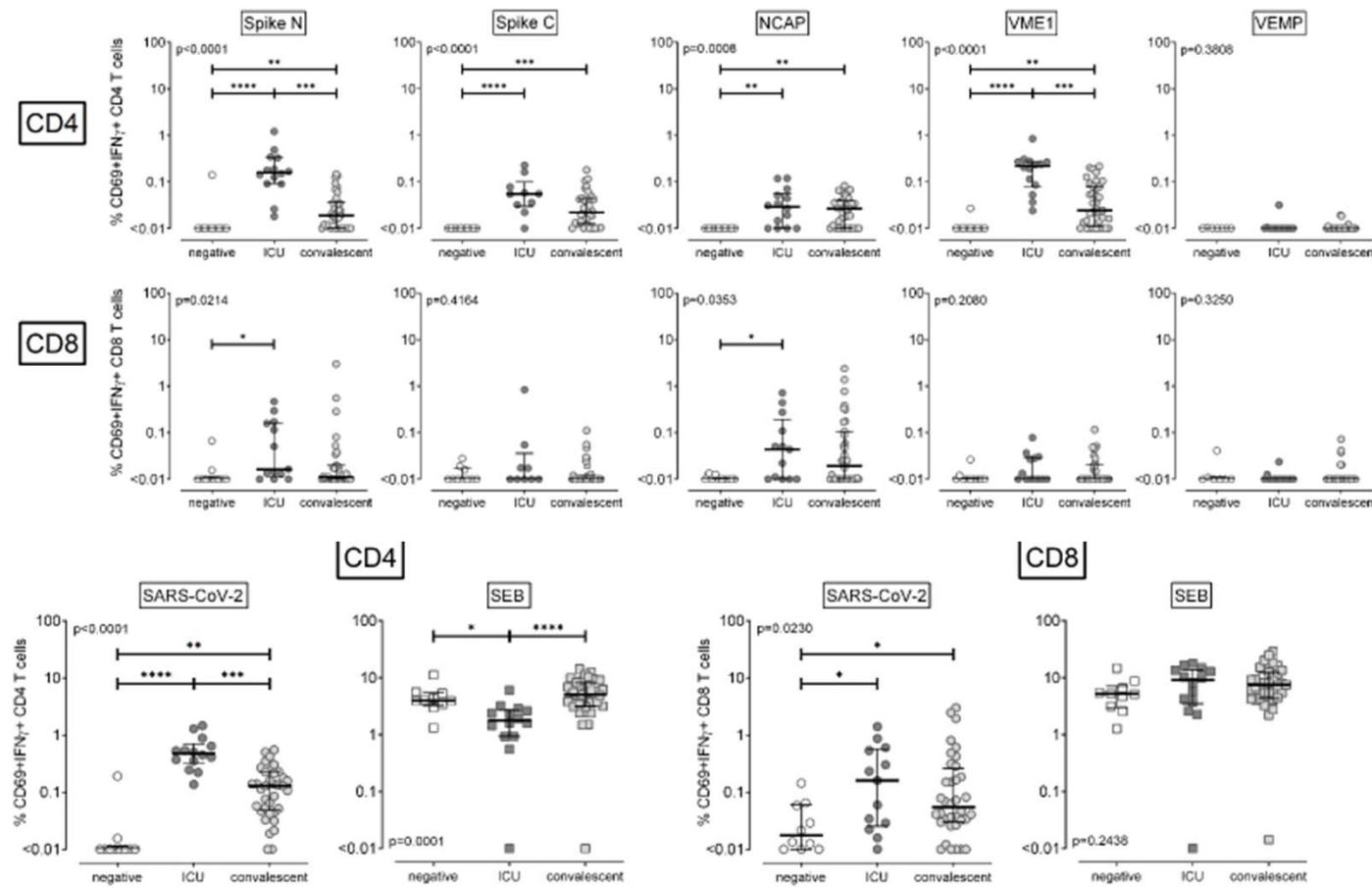


Recovering group (R); Severe persistence group (S)

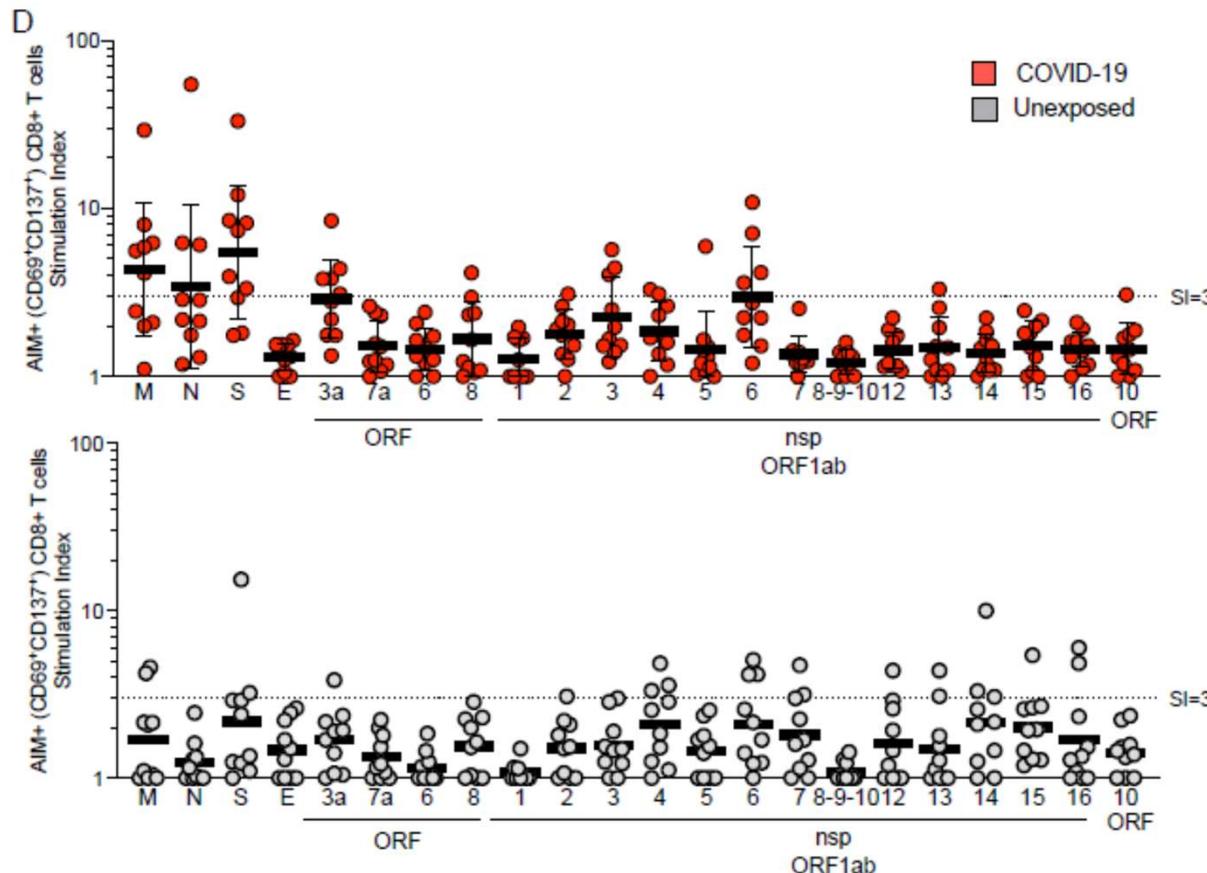
**(B) Comparison of absolute numbers of CD38+HLA-DR+CD8+ T cells (I), CD38+HLA-DR+CD4+ T cells (II) and nAbs (III) in 1 ml blood samples.**  
The data are presented as the mean  $\pm$  SEM (18 measurements from the 6 patients in R group and 9 measurements from the 5 patients excluding patient S6 in S group)

Wang et al, AJRCCM 2020

# Peripheral CD8 and mostly CD4 SARS-CoV2 –specific T cell responses are detectable during severe COVID-19 infection



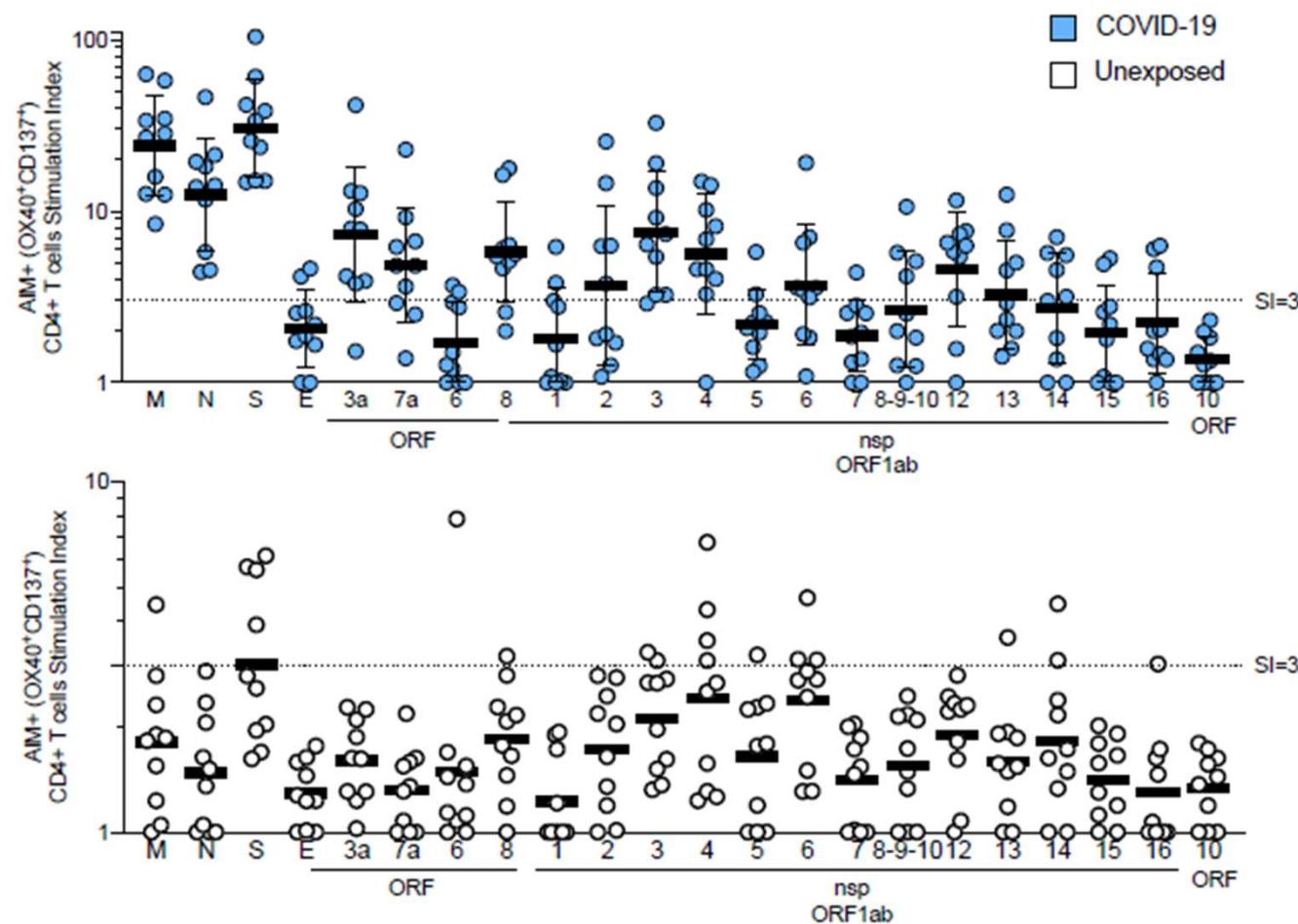
# Peripheral CD8 SARS-CoV2 –specific T cell responses are detectable after recovery from COVID-19 infection



20 adult patients who had recovered from COVID-19 disease

Disease Severity  
Mild 70% (14/20)  
Moderate 20% (4/20)  
Severe 10% (2/20)  
Critical 0% (0/20)

# Peripheral CD4 SARS-CoV2 –specific T cell responses are detectable after recovery from COVID-19 infection



20 adult patients who  
had recovered from  
COVID-19 disease

Grifoni et al, Cell 2020

# Peripheral SARS-CoV2 –specific T cell responses are detectable after mild and mostly severe COVID-19 infection

Figure 1A

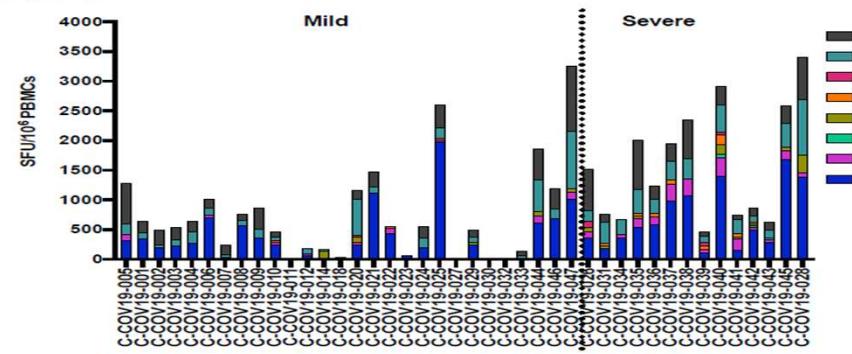
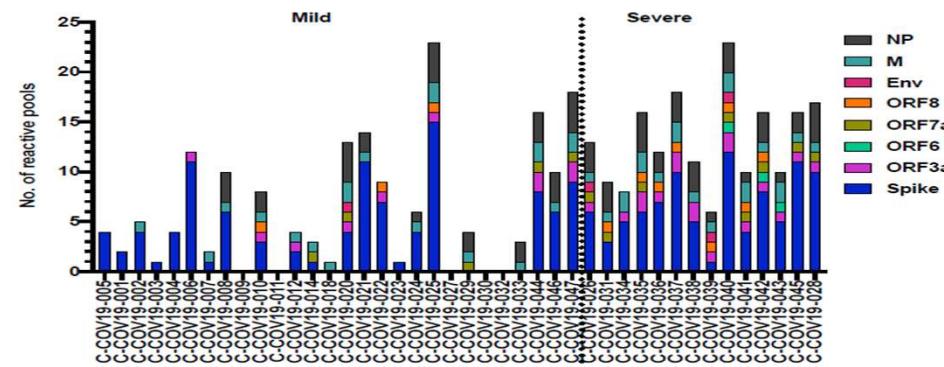
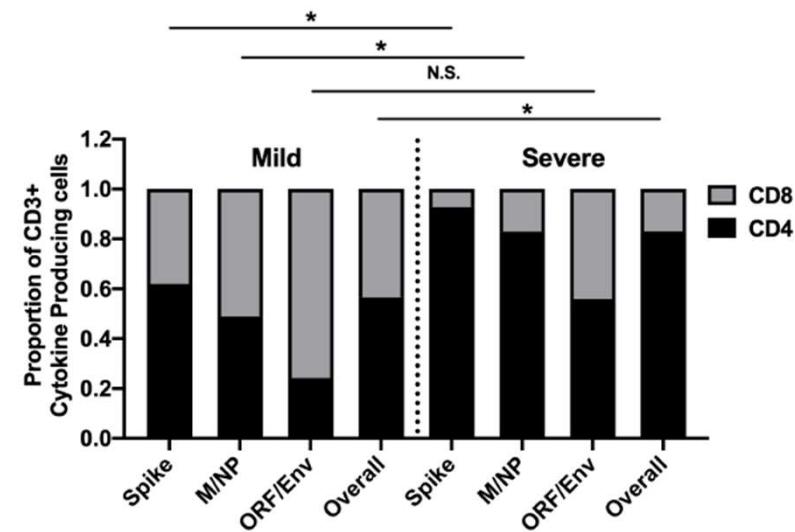


Figure 1B



A total of 423 15- to 18-mer peptides overlapping by 10 amino acid residues and spanning the full proteome of the SARS-CoV-2 except ORF-1 (253 spike, 29 M, 9 E, 35 ORF3a, 7ORF6, 15 ORF7a, 16 ORF8, 59 NP)



**Figure 4: Distribution of SARS-CoV-2-specific CD4<sup>+</sup> and CD8<sup>+</sup> memory T cell responses**

Cytokine producing T cells were detected by intracellular cytokine staining (ICS) after incubation with SARS-CoV-2 peptides. A) and B) FACS plots representative of CD4<sup>+</sup> T cell and CD8<sup>+</sup> T cells response respectively upon stimulation with respective SARS-CoV-2 peptide pools. C) The relative proportion of SARS-CoV-2 peptide pool-reactive CD4<sup>+</sup> and CD8<sup>+</sup> T cells. The SARS-CoV-2 peptide pool-reactive CD4<sup>+</sup> or CD8<sup>+</sup> T cells were identified with at least one of the three cytokines detected: IFN- $\gamma$ , TNF- $\alpha$  and IL-2. Data shown are from 14 subjects with previously mild COVID-19 symptoms and 8 with severe symptoms. Mann-Whitney test was used for the analysis. \* P<0.05

Peng et al, Nat Immunol 2020

# Local pulmonary viral replication during severe forms leads to systemic replication and immune cell recruitment

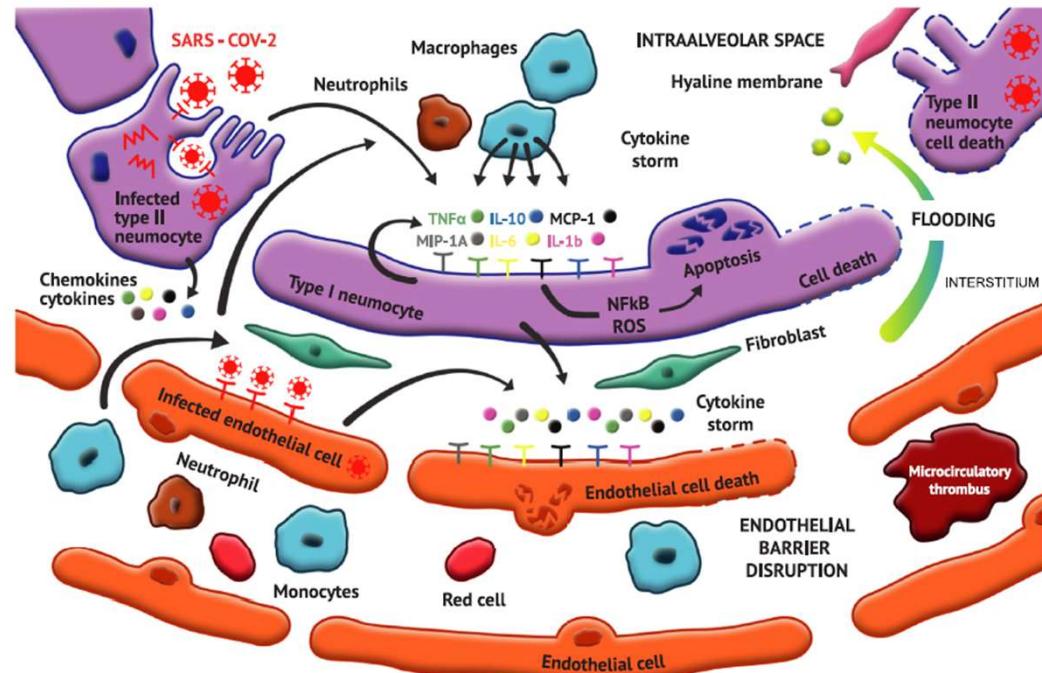
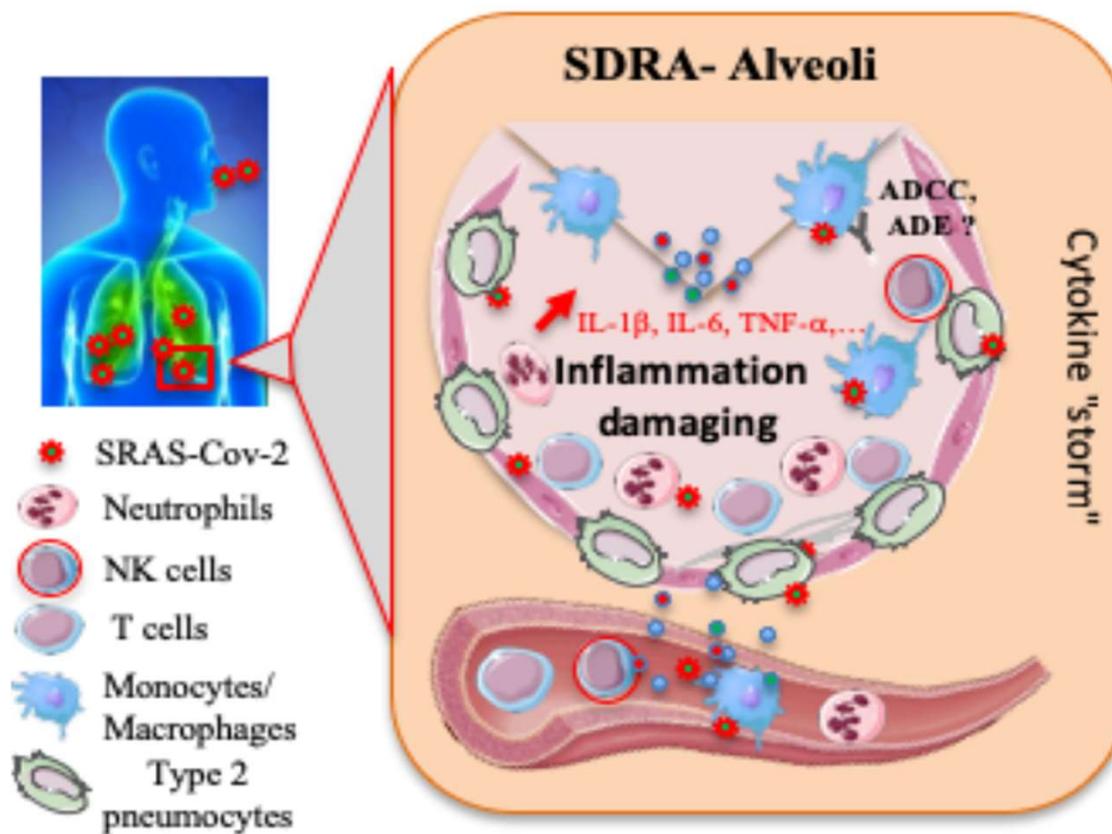


Fig. 2. Physiopathology of acute lung injury in SARS-CoV-2 infection (COVID-19).

SARS-CoV-2 infects primarily type II pneumocytes through binding to the ACE2 receptor. The infected and surrounding pneumocytes secrete cytokine and chemokines, which attract monocyte-macrophages and neutrophils to the alveolar space, which secrete additional cytokines and chemokines. Ultimately the pneumocytes suffer apoptosis/pyroptosis releasing large amounts of proinflammatory factors. Endothelial cells are infected, overexpress adhesion molecules, and release chemokines and cytokines. Endothelial cells undergo apoptosis, which, together with alveolar cell apoptosis, increases vascular leakage and breaks the alveolar-capillary barrier. The hyperinflammatory milieu and endothelial dysfunction activate coagulation cascades through tissue factor expression, platelet activation, and NETosis all of them promoting microcirculatory thrombi formation. The break of endothelial-alveolar barrier further promotes vascular leakage resulting in interstitial and alveolar space flooding. Downregulation of the ACE2/Ang-(1–7)/Mas1R axis contributes to increasing vasoconstriction, inflammatory signals, endothelial dysfunction, vascular leakage, and prothrombotic state.

Local pulmonary viral replication during severe forms leads to systemic inflammation and immune cell recruitment



Guibot et al, *Frontiers Immunol* 2020

## Conclusion : Réponses cellulaires T au cours des formes sévères de COVID-19

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- Lymphopénie T/B/NK prédominant sur les CD4 réversible après résolution de la maladie malgré une activation T persistante
  - Les cellules NK sont activées mais leur fonction est altérée
  - Présence d'une réponse T spécifique du SARS-COV-2 systémique
  - Avec migration des effecteurs au site de l'infection
  - La réponse cellulaire T est plutôt CD4 localement et en périphérie
- Les mécanismes immunopathologiques des formes létales restent à déterminer

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